

2. EXPERIMENT

We prepared poly (n-hexyl L-glutamate) (PHeLG₁₁₄) and poly (γ -benzyl L-glutamate) containing biotin group at the terminal (PBLG₂₁-bio). The polymerization degree of PHeLG₁₁₄ and PBLG₂₁-bio was estimated by ¹H-NMR and gel permeation chromatography (GPC), respectively.

Silicon wafer (Nilaco) was used as substrate. The surface of silicon is known to be hydrophilic owing to the hydroxyl groups. So the silicon was modified with silane-coupling agent to produce the hydrophobic surface [9, 10]. First, substrate was cleaned by acetone and nitric acid, and then immersed in chloroform solution of octadecyltrimethoxysilane (1mM) for 24h. And then it was kept at 110 °C for 20 min.

The surface pressure-area (π -A) measurements of the polypeptide monolayer and the monolayer deposition were performed with LB film deposition apparatus NL-BIO40-MWCT (Nippon Laser & Electronics Lab.). PHeLG₁₁₄ (chloroform/DMF(7:3) solution) and PBLG₂₁-bio (chloroform solution) were spread on a Langmuir trough filled with pure water, respectively. The deposition of these monolayer was carried out at constant surface pressure, 15mN/m for PHeLG₁₁₄ and 9.5mN/m for PBLG₂₁-bio.

Reflection spectrum measurements were performed by use of a UV/VIS spectrophotometer V-550 (JASCO) together with an attachment, ARV-474 (JASCO).

The experiment of solvent sorption on PHeLG₁₁₄ LB film was measured in 1,4-dioxane/water solution using the UV/VIS spectrometer described above. PBLG₂₁-bio was soaked into aqueous solution of avidin (1.1×10^{-7} M). Antigen-antibody reaction was detected by reflection spectrum measurements. The changes in the thickness of PBLG₂₁-bio LB film with avidin adsorption were confirmed by an atomic force microscope (AFM) Nano-Scope IIIa (Digital Instruments).

3. RESULTS AND DISCUSSION

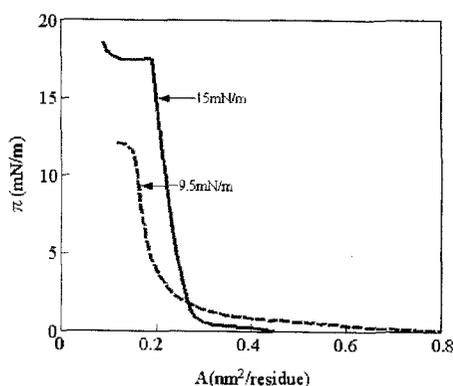


Figure 1. (π -A) isotherms for monolayers of PHeLG₁₁₄ (solid line) and PBLG₂₁-bio (dotted line).

Figure 1 shows the (π -A) isotherm of PHeLG₁₁₄ (a) and PBLG₂₁-bio (b) monolayer, respectively. Both systems formed solid state monolayer at air/water interface. The limiting area was estimated to be 0.26nm² / residue (PHeLG₁₁₄) and 0.21nm² / residue (PBLG₂₁-bio), respectively. This value indicated that α -helix rod of PHeLG₁₁₄ and PBLG₂₁-bio are lying on the air/water interface.

To create structural color substrates, we tried to transfer the several polypeptides monolayers onto the silicon substrates. PHeLG₁₁₄ was deposited at the surface pressure of 15mN/m. PHeLG₁₁₄ LB film showed structural color depending on the number of deposition, blue (80 layers), yellow (120 layers), red (160 layers). On the other hand, PBLG₂₁-bio monolayer was deposited at the surface pressure of 9.5mN/m. In this case, silicon substrate was annealed at 1000°C for 3h before use to get a pre-colored substrate. 10 layers of PBLG₂₁-bio film on this pre-colored substrate showed viridian color. These structural color substrates change their color depending on the incident angle of light.

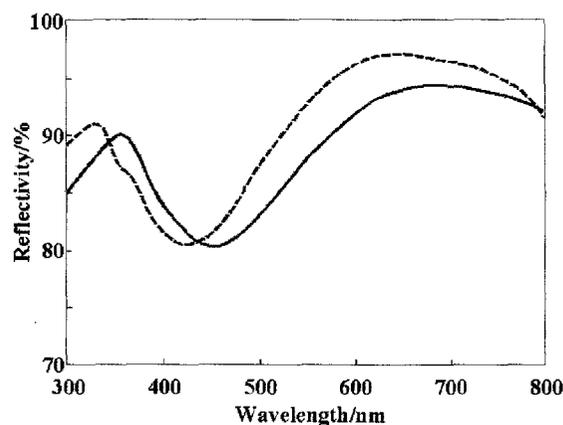


Figure 2. Reflective UV/VIS spectra of PHeLG₁₁₄ LB film (120 layers) in water (dotted line) and in 1.95M 1,4-dioxane/water solution (solid line).

These colored LB films can be characterized by their reflection spectra of LB film. Figure 2 shows reflective UV/VIS spectra of 120 layers of PHeLG₁₁₄ LB film in pure water and in 1.95M 1,4-dioxane/water solution at an incident angle of 10°. As a result of the transfer of PHeLG₁₁₄ LB film from pure water to 1,4-dioxane/water solution, maximum peak was red-shifted from 634nm to 687nm. 1,4-dioxane is good solvent for the polypeptide. It was prospective that the sorption of 1,4-dioxane caused swelling of PHeLG₁₁₄ LB film. The difference of refractive index between polypeptide ($n=1.5$) and 1,4-dioxane ($n=1.42$) is very small. So the red-shift may be owing to the solvent induced increase in the thickness

of LB film.

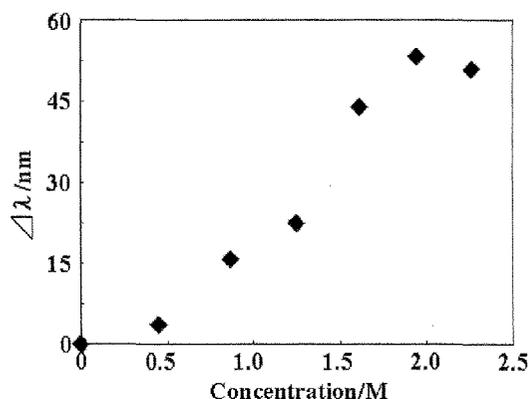


Figure 3. Changing in $\Delta\lambda$ of PHeLG₁₁₄ LB film (120 layers) with 1,4-dioxane concentration in water.

Figure 3 shows the degree of red-shift, $\Delta\lambda$, dependence on the concentration of 1,4-dioxane. The value of $\Delta\lambda$ monotonously increased with increasing the concentration of 1,4-dioxane. This implies a possibility for visual sensing of the concentration of chemicals in solution. It was also confirmed that sorption of 1,4-dioxane vapor on PHeLG₁₁₄ LB film drastically changed the structural color. It should be noted that this color change occurred homogeneously over the whole area of the LB film.

It was confirmed, therefore that the sorption of solvent causes structural color change. However, it is difficult to specifically detect a chemical among the mixture. So as a next step, we tried to construct the visual sensing system for antigen-antibody reaction.

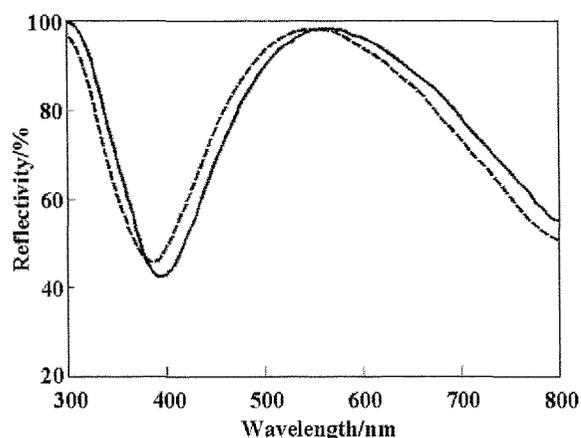


Figure 4. Reflective UV/VIS spectra of SiO₂ layer with 10 layers of PBLG₂₁-bio (dotted line) and after soaking avidin/water solution (solid line).

Figure 4 shows reflective UV-VIS spectra of PBLG₂₁-bio LB film (10 layers) on the pre-colored substrate before and after the soaking into avidin/water solution ($1.1 \times 10^{-7} M$ 3h) at an incident angle of 10°. It is clear that the maximum peak of the spectra was shifted 20nm longer wavelength. Eq.(1) shows that 20nm red-shift corresponds to 7.2nm increasing in the thickness of the LB film.

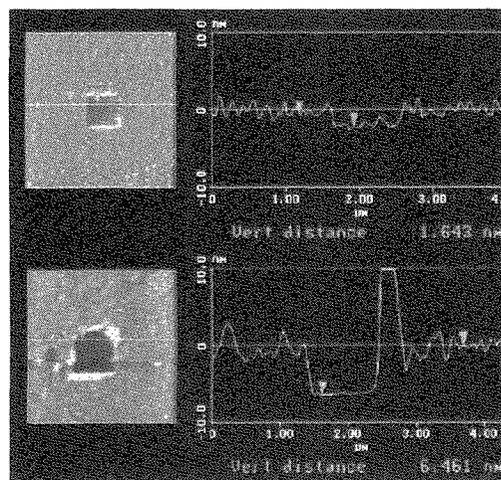


Figure 5. AFM images of 1 layer of PBLG₂₁-bio LB film and after soaking avidin/water solution on Si/SiO₂ surface.

AFM images (Figure 5) show the change in the thickness of PBLG₂₁-bio LB film before and after soaking avidin/water solution. The increasing in the thickness was estimated to be about 5nm. Considering that the diameter of avidin is about 6nm, it was expected avidin adsorbed on the biotin site on the surface of PBLG₂₁-bio LB film. This may be a reason for the structural color change. This result also shows the possibility of visual sensing of antigen-antibody reaction.

4. CONCLUSION

In this study, we succeeded in preparation of structural color LB films with α -helical hydrophobic polypeptide and α -helical hydrophobic polypeptide containing biotin group on the silicon substrate. They showed various interference colors depending on the number of layers. The structural color could be changed by sorption of 1,4-dioxane which caused swelling of PHeLG₁₁₄ LB film. This red-shift was dependent on the 1,4-dioxane concentration. On the other hand, the substrate prepared by using α -helical polypeptide containing biotin group, changed its structural color by adsorption of avidin

owing to the increasing in the thickness. This shows a possibility of visual sensing for antigen-antibody reaction.

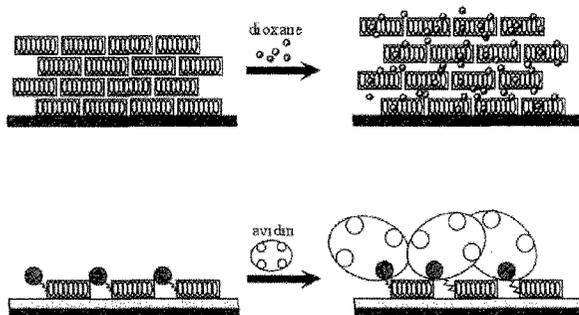


Figure 6. Systems of changing structural color caused by 1,4-dioxane sorption and avidin adsorption.

The results obtained in this study may be applicable to novel visual sensing systems for the host-guest chemistry.

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